

# 1 Construction of compostable packaging with 2 antibacterial property and improved performance using 3 sprayed coatings of modified cellulose nanocrystals

4 Shuting Huang<sup>1</sup>, Sheng Zou<sup>1</sup>, Yixiang Wang<sup>1, \*</sup>

5 <sup>1</sup> Department of Food Science and Agricultural Chemistry, McGill University, Ste Anne  
6 de Bellevue, Quebec, Canada H9X 3V9

7 \* Correspondence: [yixiang.wang@mcgill.ca](mailto:yixiang.wang@mcgill.ca), +001(514)3987922

## 8 Abstract

9 Increasing concerns about food safety and the environment have facilitated the  
10 development of eco-friendly antibacterial packaging. This study aimed to demonstrate a  
11 facile way to fabricate active packaging materials with modified cellulose nanocrystals  
12 (CNCs) and compare the effects of different modified CNCs on the performance of  
13 compostable materials. Polylactic acid (PLA) film was selected as a model, and CNCs were  
14 modified with methacrylamide, cetyltrimethylammonium bromide, and zinc oxide,  
15 respectively, and then applied on the surface of PLA films by spray-coating. All modified  
16 CNCs showed excellent antibacterial activity against *S. aureus* and *E. coli* (>99.999%).  
17 The effects of different CNC modifications on the performance of PLA films were  
18 investigated. Compared to neat PLA films, PLA/CNC films exhibited improved  
19 mechanical strength with maintained flexibility, lower gas permeability, and faster compost  
20 disintegration rate, and extended the shelf life of wrapped pork samples from 3 days to  
21 more than 10 days. Therefore, this work will also facilitate the applications of PLA  
22 materials in eco-friendly packaging.

23 **Keywords:** cellulose nanocrystals, modifications, polylactic acid, active packaging,  
24 antibacterial activity, compost disintegrability

## 25 **1. Introduction**

26 Foodborne diseases are global public health concerns that cause considerable  
27 socioeconomic implications. In fact, the problem is caused by consuming food  
28 contaminated with microorganisms or chemical substances that result in over 200 diseases  
29 (Gao et al., 2022). According to the estimation of the World Health Organization,  
30 foodborne diseases lead to over 420,000 deaths annually, with 40% being children under  
31 five (WHO, 2020). Among the solutions to reduce foodborne diseases, antibacterial  
32 packaging is an important approach to controlling foodborne pathogens during  
33 transportation and storage. At the same time, the increasing environmental concerns about  
34 non-degradable plastics bring awareness of “green” packaging materials. Numerous  
35 research efforts have been made to promote the applications of biodegradable polymers,  
36 which include polysaccharides, proteins of animal or plant origin, lipids, and polyesters  
37 from microbial sources (Fonseca-García et al., 2021; Omerović et al., 2021). Among them,  
38 polylactic acid (PLA) has been reported as a promising candidate to replace the fossil-  
39 based plastic, due to its renewability, biodegradability, facial processing, and  
40 biocompatibility (Ma et al., 2022; Shojaeiarani et al., 2022). PLA has the largest market  
41 share in the biodegradable plastic field, estimated for a market of \$6.5 billion by 2025  
42 (Shojaeiarani et al, 2020). Various PLA based antimicrobial packaging materials have been  
43 developed by the incorporation of functional agents such as metal oxide (Zhang et al.,  
44 2021), lignin (Cerro et al., 2021), chitosan (Kongkaoroptham et al., 2021), and essential oil  
45 (Fiore et al, 2021). For instance, Jiang et al. (Jiang et al., 2022) prepared antibacterial  
46 membranes based on PLA, polybutylene adipate terephthalate, carboxymethyl cellulose,  
47 and silver, which showed the effective antibacterial property against *Staphylococcus*  
48 *aureus* (*S. aureus*) and *Escherichia coli* (*E. coli*). However, the poor mechanical and barrier  
49 properties of PLA films limited their applications in food packaging due to the low  
50 crystallinity of PLA (Beauson et al., 2022; Sharafi et al., 2022).

51 Cellulose nanocrystals (CNCs) have received noticeable attention owing to their unique  
52 morphology, high mechanical strength, and broad functionalization capacities (Huang et

53 al., 2020). Numerous studies focused on the reinforcing effects of CNCs on the mechanical  
54 properties of packaging films (Niinivaara et al., 2021; Rojas-Lema et al., 2021; Salmieri et  
55 al., 2014a, 2014b). To further broaden the application scope of CNCs, recent research  
56 investigated the modifications of CNCs with multifunctions and applied them as functional  
57 nanofillers of packaging materials. For example, Koshani et al. (Koshani et al., 2021)  
58 synthesized the antibacterial hairy CNCs with the conjugated photosensitizer rose bengal,  
59 which could inactivate over 80% of both *Listeria monocytogenes* and *Salmonella enterica*  
60 serotype Typhimurium upon normal light irradiation. Shin et al. (Shin et al., 2022) prepared  
61 the functionalized CNCs by grafting 3-pentadecylphenol that showed the antibacterial  
62 property of 99.99% against *E. coli*. However, these nanofillers needed to be incorporated  
63 when the packaging materials were formed, and the effects of different modified CNCs on  
64 compostable packaging have been seldom compared.

65 Packaging materials based on biopolymers will help reduce the negative effect on the  
66 environment, while the antibacterial property can better protect food products and decrease  
67 food waste. In this study, we hypothesized that (i) a sprayed coating of modified CNCs  
68 could improve the performance of PLA films, and (ii) the effects were related to the  
69 different modifications of CNCs. CNCs derived from waste textile were selected as a model  
70 and modified with N-halamine (methacrylamide, MAM), quaternary ammonium salt  
71 (CTAB), and metal oxide (ZnO), respectively. The modified CNCs were spray-coated on  
72 the surface of PLA films, and their effects on the properties (antibacterial activity,  
73 mechanical properties, gas barrier properties, and compost disintegrability) of PLA films  
74 were investigated. Moreover, the preservation of pork loin with CNC-coated or uncoated  
75 PLA films was monitored.

## 76 **2. Materials and Methods**

### 77 **2.1. Materials**

78 Waste cotton clothes were kindly provided by Renaissance (Montreal, QC, Canada). CNCs  
79 were directly extracted from textile waste by using sulfuric acid hydrolysis, according to  
80 our previous work (Huang et al., 2020). Cetyltrimethylammonium bromide (CTAB,  $\geq 98\%$ ),

81 zinc acetate dihydrate (>95%), sodium hydroxide (NaOH,  $\geq 97.0\%$ ), methacrylamide  
82 (MAM, 98%), sodium persulfate ( $\geq 98\%$ ), sodium thiosulfate anhydrous ( $\geq 98\%$ ),  
83 tetrahydrofuran (THF, >95%), and glacial acetic acid ( $\geq 99.7\%$  w/w) were purchased from  
84 Fisher Scientific (Ottawa, ON, Canada) and used without further treatment. Pellet PLA  
85 (Ingeo 4043D grade, >98%, density of 1.24 g/cc) was purchased from NatureWorks  
86 (Minnetonka, MN, USA). Luria–Bertani (LB) medium and tryptic soy agar (TSA) were  
87 purchased from Becton, Dickinson and Company (Franklin Lakes, NJ, USA), and  
88 phosphate-buffered saline (10 $\times$  PBS) was obtained from VWR International (Mississauga,  
89 ON, Canada).

## 90 **2.2. Modification of CNCs**

91 MAM-modified CNCs were prepared according to the method reported by Liu et al. (Liu  
92 et al., 2017) with some modifications. The desired amount of NaOH was added into 50.00  
93 g of CNC aqueous suspension (10 wt%), and then 5.00 g MAM and 0.03 g sodium  
94 persulfate were added and stirred at room temperature for 20 min. The reaction was  
95 conducted at 65 °C (in a water bath) for 5 hours. After that, the product was washed with  
96 distilled water to remove unreacted MAM and sodium persulfate and coded as MAM-  
97 CNCs.

98 The preparation of CTAB-modified CNCs was carried out according to the method  
99 described by Ranjbar et al. (Ranjbar et al., 2020). Briefly, 40.00 g of CNC aqueous  
100 suspension (5 wt%) was slowly added to 20.00 g of CTAB solution (5 wt%). The mixture  
101 was stirred at room temperature for 2 hours. Then, the modified CNCs were washed with  
102 distilled water to remove the unbounded CTAB and coded as CTAB-CNCs.

103 The zinc oxide-modified CNCs were synthesized by using zinc acetate as the zinc precursor  
104 and NaOH as a reducing agent, according to a previous report (Badawy et al., 2021) with  
105 some modifications. In brief, zinc acetate solution (50.00 g, 10 wt%) was mixed with CNC  
106 aqueous suspension (50.00 g, 2.5 wt%) and stirred at 80 °C for 1 hour. Then, NaOH solution  
107 (50 mL, 0.1 mol/L) was added dropwise to the mixture, and the suspension showed a milky  
108 color. The reaction continued at 80 °C for 2 hours under stirring. The obtained samples

109 were well-washed with distilled water and coded as ZnO-CNCs.

### 110 **2.3. Preparation of PLA/CNC composite films**

111 PLA films were prepared by dissolving 0.8 g PLA pellets in 10 mL THF, and the solution  
112 was filled into a glass mold and dried at 25 °C for 12 hours. After washing with distilled  
113 water, the modified CNCs were centrifugated (7000 g, 30 min, Eppendorf centrifuge 5430,  
114 NRW, Germany), dispersed in acetic acid (2 w/v%), and then sprayed on the surface of  
115 PLA films (length × width × thickness: 8 cm × 6 cm × 0.15 mm). The PLA/CNC composite  
116 films were dried in the fume hood for 24 hours and coded as PLA-CC4, PLA-CC8, PLA-  
117 ZC4, PLA-ZC8, PLA-MC4, and PLA-MC8, corresponding to the CTAB-CNCs, ZnO-  
118 CNCs, and MAM-CNCs contents (based on the dry weight of PLA) of 4 and 8 wt%,  
119 respectively. The films were stored at 25 °C and 50% relative humidity for 3 days prior to  
120 analysis.

### 121 **2.4. Characterizations**

#### 122 **2.4.1. Fourier-transform infrared (FTIR) spectroscopy**

123 FTIR spectra of modified CNCs and PLA/CNC composite films were obtained by using a  
124 Cary 630 FTIR spectrometer with an attenuated total reflectance sampling module (Agilent  
125 technologies, Inc., USA). The spectra were collected in the range of 4000-650 cm<sup>-1</sup> as the  
126 average of 64 scans with a resolution of 2 cm<sup>-1</sup>, using the empty accessory as blank.

#### 127 **2.4.2. X-ray diffraction (XRD)**

128 XRD patterns of pristine and modified CNCs were obtained using an Empyrean 3 (Malvern  
129 Panalytical Ltd., UK) X-ray diffractometer in a Bragg Brentano configuration, with Cu K $\alpha$   
130 radiation between 4° and 80°. The crystallinity index (CrI) was determined by the peak  
131 height method (Park et al., 2010) in terms of Equation (1):

$$132 \text{CrI (\%)} = \frac{I_{(200)} - I_{am}}{I_{(200)}} \times 100\% \quad \text{Equation (1)}$$

133 where  $I_{(200)}$  is the maximum diffraction intensity associated with surface areas of crystalline  
134 cellulose, and  $I_{am}$  is the diffraction intensity of an amorphous cellulose fraction.

#### 135 **2.4.3. UV-vis spectroscopy**

136 UV-vis spectra of pristine CNC and ZnO-CNC suspensions were collected on a DU 800

137 UV-Vis spectrophotometer (Beckman Coulter, USA) in the wavelength range of 200-400  
138 nm against distilled water as blank.

#### 139 **2.4.4. Zeta potential measurement**

140 The surface charge of CNC samples was measured using the NanoBrook Omni zeta  
141 potential analyzer (Brookhaven Instruments Corporation, USA). Triplicate measurements  
142 were taken at 25 °C after the samples were conditioned for 300 seconds.

#### 143 **2.4.5. Transmission electron microscopy (TEM)**

144 The morphology of modified CNCs was observed using the Talos F200X G2 TEM  
145 (Thermo Fisher Scientific, USA). A tiny drop of diluted CNC suspension was deposited on  
146 a carbon-coated copper grid. After air drying at room temperature, the sample was imaged  
147 on TEM at a voltage of 200 kV.

#### 148 **2.4.6. Scanning electron microscopy (SEM)**

149 The surface morphology of PLA/CNC composite films was observed by Hitachi TM1000  
150 SEM (NJ, USA), operating at an acceleration voltage of 4 kV. The film samples were  
151 sputtered with 4 nm gold-platinum prior to observation and photographing.

#### 152 **2.4.7. Antibacterial test**

153 Two strains were used for the antibacterial testing, namely *S. aureus* ATCC 6538 (Gram-  
154 positive) and *E. coli* K12 (Gram-negative). Each strain was prepared from -80 °C 20%  
155 glycerol stock and streaked for isolation on LB agar plates. The plates were incubated at  
156 37 °C overnight, and isolated colonies were picked for inoculation of 4 mL LB broth at  
157 37 °C with constant shaking. After 16 hours, a concentration of ca. 8 log for each bacterial  
158 culture was achieved, and the bacterial inoculums were prepared by diluting the culture  
159 with PBS to a certain concentration for further experiments.

160 The antibacterial efficacy of modified CNCs was evaluated by mixing 20 mL of CNC  
161 suspension and 200 µL of bacterial inoculum to a final concentration of 10<sup>5</sup> CFU/mL (Tang  
162 et al., 2020). The mixture was incubated for 1 hour with constant shaking, and then the  
163 upper suspension was used for serial dilution using PBS. After that, the diluted solution  
164 was spread onto the surface of LB agar plate and incubated at 37 °C for 24 hours. The

165 colonies on the plate were enumerated to calculate the log reduction.  
166 The antibacterial activity of PLA/CNC composite films against *S. aureus* and *E. coli* was  
167 analyzed by a modified AATCC 100 test method (Xu et al., 2021). The composite films  
168 with a thickness of 0.15 mm were cut into 2 cm × 2 cm. Then, 20 μL of bacterial suspension  
169 (~10<sup>6</sup> CFU/mL) was sandwiched between two films to enable sufficient contact. After 1  
170 hour of contact, 4 mL of sterile sodium thiosulfate was added and vortexed to rinse off  
171 bacteria. The rinsing solution was serially diluted for the plating assay.

172 The zone of inhibition was measured using the disc diffusion method (Zhang et al., 2021).  
173 *S. aureus* and *E. coli* suspensions with a concentration of around 10<sup>8</sup> CFU/mL were evenly  
174 spread on the LB agar plates, and the composite films (diameter of 6 mm) were placed over  
175 the surface of the inoculated plate. The plates were incubated at 37 °C for 24 hours, and  
176 the bacterial inhibition zone was measured.

#### 177 **2.4.8. Mechanical properties**

178 The mechanical properties of PLA/CNC composite films were tested using an eXpert 7601  
179 single column testing machine (ADMIT, USA) at 25 °C according to the standard ASTM  
180 D882. The dimension of film specimens was 60 mm × 10 mm × 0.15 mm (length × width  
181 × thickness). The initial grip separation distance was set as 20 mm, and the separation speed  
182 was 20 mm/min. The thickness of the films was measured by a Traceable digital caliper  
183 (Fisher Scientific, ON, Canada).

#### 184 **2.4.9. Gas barrier properties**

185 Water vapor permeability (WVP) was determined using a water vapor permeability tester  
186 (model 3/61, Mocon, Inc., USA) at 37 °C and 90% relative humidity (RH). WVP values  
187 were calculated using Equation (2) (Zhou et al., 2022):

$$188 \quad WVP = \frac{WVTR \times n}{\Delta p} \quad \text{Equation (2)}$$

189 where  $n$  is the film thickness (m), and  $\Delta p$  is the partial pressure difference across the films  
190 (Pa).

191 The oxygen transmission rate (OTR) of PLA/CNC composite films was determined at 23 °C

192 and 0% RH using an oxygen permeability tester (model 2/22, Mocon, Inc., USA).

#### 193 **2.4.10. Water contact angle**

194 The water contact angle was tested by a contact angle meter (Future digital scientific, Co.  
195 USA) using the sessile drop method. The water droplet (5  $\mu$ L) was placed on the film  
196 surface, and the image was immediately captured and analyzed with the goniometer at  
197 room temperature. Each measurement was performed on a different spot of the film, and  
198 the results were based on the average of three measurements.

#### 199 **2.4.11. Disintegration under composting conditions**

200 The disintegration performance of PLA/CNC composite films was tested following the  
201 ISO-20200 standard (ISO, 2015). Briefly, solid synthetic waste was prepared by mixing  
202 40% sawdust, 30% rabbit feed, 10% ripe compost, 10% corn starch, 5% saccharose, 4%  
203 corn oil, and 1% urea together. After that, the dry waste was mixed with water in 45:55  
204 ratio. The composite films were cut into 25 mm  $\times$  25 mm and buried at 6 cm depth in the  
205 composting reactor containing reconstituted wet waste. The reactor was then put in an air-  
206 circulation oven at  $58 \pm 2$  °C, and water was added periodically to maintain the humidity  
207 in the compost. The films were recovered from the reactor at different times (3, 7, 10, and  
208 14 days), dried, and weighed. The weight loss of the films was calculated using Equation  
209 (3) (Cerro et al., 2021):

$$210 \text{ Weight loss (\%)} = \frac{m_i - m_d}{m_i} \times 100\% \quad \text{Equation (3)}$$

211 where  $m_i$  and  $m_d$  were the initial dry mass and residue dry mass of tested samples,  
212 respectively.

#### 213 **2.4.12. Meat preservation test**

214 Pork loin was selected as a food model and purchased from a local market. The pork was  
215 cut into 1 cm  $\times$  1 cm  $\times$  1 cm portions and randomly divided into five groups. The test  
216 groups were packaged with PLA/CNC composite films, and the control groups were either  
217 unpackaged or wrapped with neat PLA films. The samples were placed on the trays and  
218 stored at 4 °C. The total viable count (TVC) values were determined at 0, 1, 3, 5, 7, and 10  
219 days of storage according to the method reported by Zhong et al. (Zhong et al., 2021).



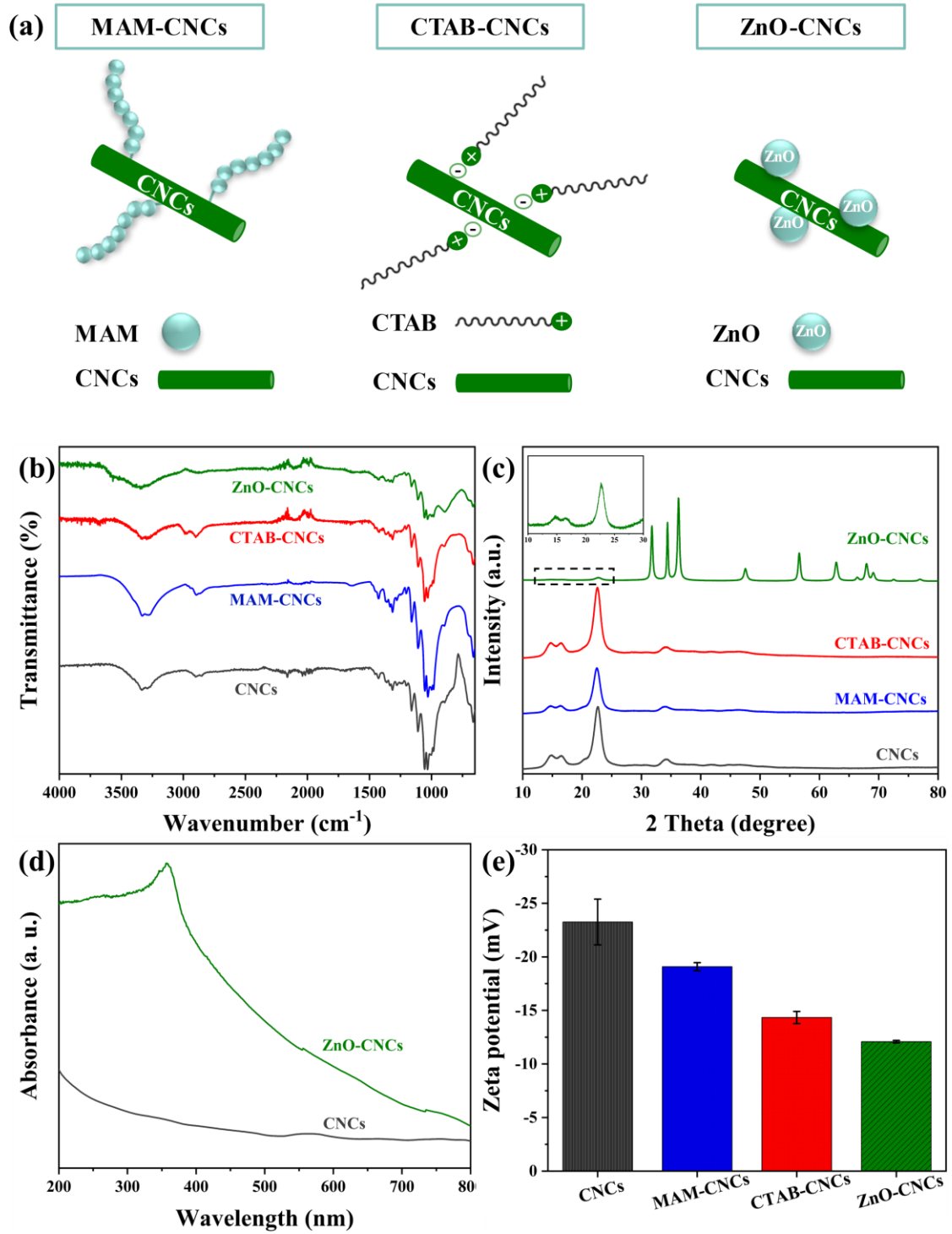
220 Briefly, the pork samples were transferred aseptically to a stomacher bag and added with  
221 PBS solution. The mixture was homogenized for 2 minutes using a lab stomacher blender  
222 (Seward, UK). Then, serial dilutions were prepared with PBS, spread on TSA plates, and  
223 incubated at 37 °C for 48 hours. The experiments were performed in triplicate, and the  
224 results of TVC were reported as CFU/g.

## 225 **2.5. Statistical analysis**

226 The experiments were carried out in triplicate, and data were presented as the mean  $\pm$   
227 standard deviation. The statistical analysis of the data was carried out through a one-way  
228 analysis of variance (ANOVA) using IBM SPSS Statistics 26 software, and the differences  
229 between means were analyzed by LSD post-hoc analysis at the confidence level of 0.05.

230 **3. Results and Discussion**

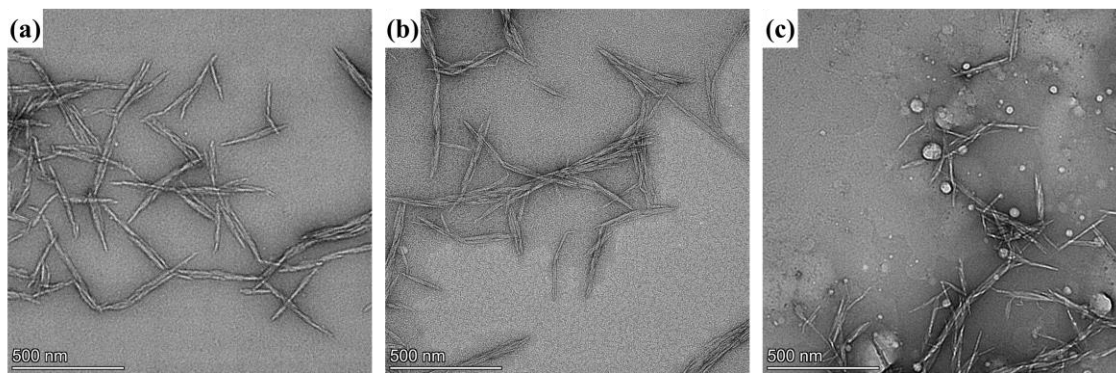
231 **3.1. Structure of modified CNCs**



232

233 **Figure 1.** (a) Illustration, (b) FTIR spectra, (c) XRD patterns, (d) UV-vis spectra, and (e)

234 zeta potential of pristine and modified CNCs.

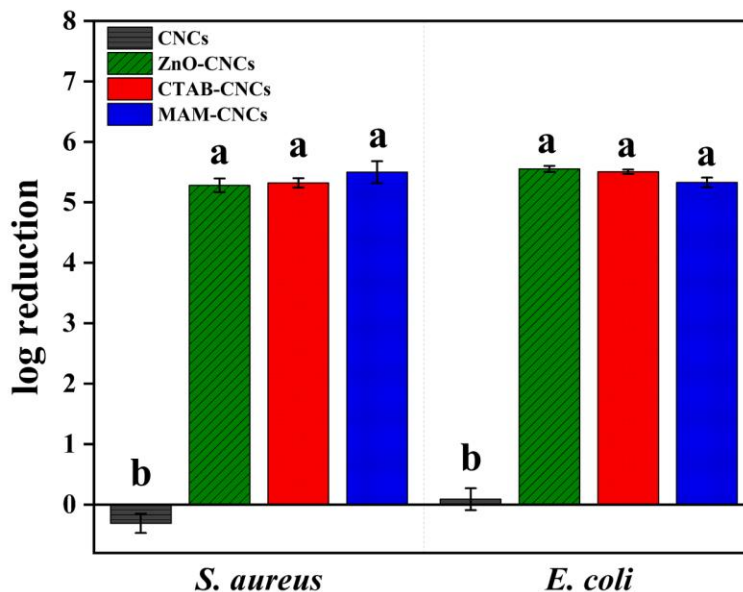


**Figure 2.** TEM images of (a) MAM-CNCs, (b) CTAB-CNCs, and (c) ZnO-CNCs.

235  
 236  
 237 Figure 1 (a) illustrates the different modified CNCs, and their chemical structures were  
 238 analyzed by FTIR (Figure 1 (b)). The FTIR spectrum of unmodified CNCs showed the  
 239 characteristic peaks at  $3338\text{ cm}^{-1}$ ,  $2893\text{ cm}^{-1}$ ,  $1427\text{ cm}^{-1}$ ,  $1371\text{ cm}^{-1}$ ,  $1315\text{ cm}^{-1}$ ,  $1160\text{ cm}^{-1}$ ,  
 240  $1055\text{ cm}^{-1}$ ,  $1031\text{ cm}^{-1}$ , and  $893\text{ cm}^{-1}$ , corresponding to O-H stretching, C-H stretching, C-H  
 241 bending,  $\text{CH}_2$  bending, O-H bending, asymmetric vibration of C-O-C, pyranose ring  
 242 stretching, and  $\beta$ -glycoside bonds of the glucose ring, respectively. After the modification  
 243 with MAM, the FTIR spectrum of MAM-CNCs showed a significant new peak at  $1643\text{ cm}^{-1}$   
 244  $\text{cm}^{-1}$  assigned to amide carbonyl C=O bond. Besides, the peak at  $1600\text{ cm}^{-1}$  attributed to  
 245 C=C bond of MAM was not observed in MAM-CNCs, indicating that the residue of  
 246 unreacted MAM monomer was removed from the product. Proof of the successful  
 247 modification of MAM onto CNCs was interpreted in comparison with the previous report  
 248 (Rosace et al., 2017). Compared to original CNCs, only slight changes of FTIR spectrum  
 249 were observed for CTAB-CNCs. Strong C-H stretching at  $2984\text{ cm}^{-1}$  and  $2900\text{ cm}^{-1}$ ,  
 250 corresponding to the long-chain alkyl group of CTAB, suggested the presence of CTAB on  
 251 CNCs and the absence of direct chemical bonding (Ranjbar et al., 2020). For ZnO-CNCs,  
 252 no changes were found in the FTIR spectrum (Sharma et al., 2019), but a new and strong  
 253 characteristic peak at  $365\text{ nm}$  appeared in the UV-vis spectrum (Figure 1 (d)), ascribed to  
 254 the basic bandgap absorption of ZnO (Xiao et al., 2020). It has been reported that hydrogen  
 255 bonds and electrostatic attraction are two potential interactions between CNCs and ZnO  
 256 (Fu et al., 2015; Sharma et al., 2019; Zhao et al., 2017). The XRD patterns of pristine and

257 modified CNCs are shown in Figure 1 (c). All the samples exhibited similar diffraction  
258 peaks at  $14.6^\circ$  ( $1\bar{1}0$ ),  $16.4^\circ$  (110), and  $22.5^\circ$  (200), ascribed to cellulose  $I_\beta$  crystal structure  
259 (Duarte Urueña et al., 2021). ZnO-CNCs also showed the peaks at  $31.8^\circ$ ,  $34.5^\circ$ ,  $36.2^\circ$ ,  
260  $47.8^\circ$ ,  $56.5^\circ$ ,  $62.8^\circ$ , and  $68.0^\circ$ , corresponding to (100), (002), (101), (102), (110), (103),  
261 and (112) of the ZnO crystal structure, respectively (Elfeky et al., 2020). The surface  
262 charges of the modified CNCs are shown in Figure 1 (e). The original CNCs had a zeta  
263 potential value of around -23 mV, which was similar to the samples from sago frond wastes  
264 prepared by sulfuric acid hydrolysis (Arnata et al., 2020; Asadi et al., 2021). After surface  
265 modification, MAM-CNCs showed similar surface charges with CNCs. The electrostatic  
266 interaction between CTAB and CNCs resulted in the reduced negative charges of CNCs  
267 (Baggio et al., 2022), while ZnO-CNCs had the least charges due to the attracted ZnO  
268 (Badawy et al., 2021; Guan et al., 2019). Figure 2 shows the morphologies of modified  
269 CNCs, which had a typical rod-like shape with an average length of 158 nm and a diameter  
270 of 10 nm, without obvious aggregation (Badawy et al., 2021; Gahrooe et al., 2021; Li et  
271 al., 2020). The particle size of ZnO in Figure 2 (c) was  $36\pm 16$  nm, which was similar to the  
272 reported ones (Elfeky et al., 2020; Lizundia et al., 2018).

273 **3.2. Antibacterial activity**

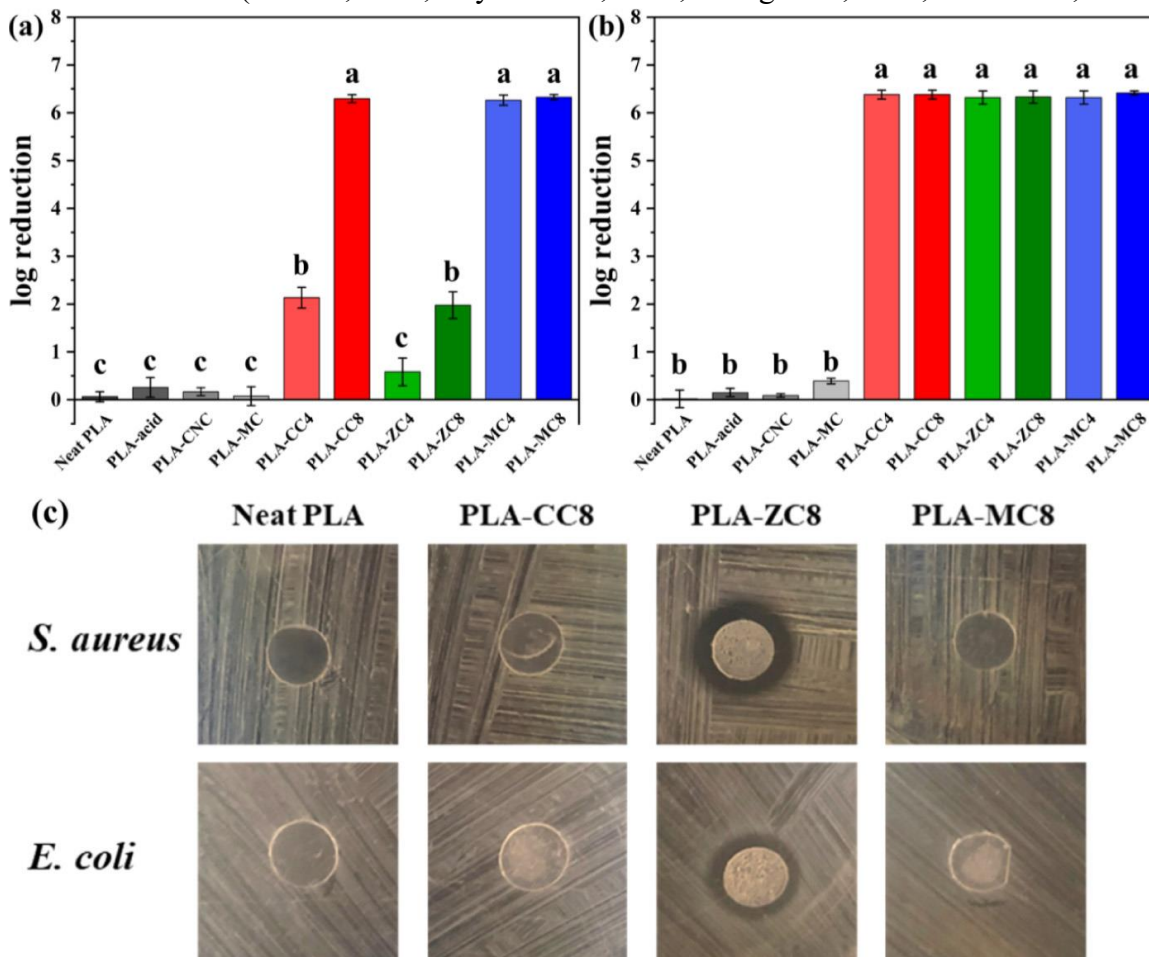


274

275 **Figure 3.** Antibacterial efficiency of pristine and modified CNCs against *S. aureus* and *E.*  
276 *coli*. Statistical significance ( $p < 0.05$ ) between different CNC samples was indicated by  
277 different letters.

278 The pristine and modified CNCs were cultivated with Gram-positive (*S. aureus*) and Gram-  
279 negative (*E. coli*) bacteria to evaluate their antibacterial activity. For MAM-CNCs,  
280 chlorination treatment with sodium hydrochloride was conducted to convert the N-H bond  
281 to oxidative chlorine, which activated the antibacterial capacity of MAM-CNCs (Chang et  
282 al., 2018). In contrast, ZnO-CNCs and CTAB-CNCs did not require any pre-treatment. As  
283 shown in Figure 3, the original CNCs did not exhibit any effect against both *S. aureus* (-  
284 0.31 log) and *E. coli* (0.09 log), which was consistent with a previous report (Zhou et al.,  
285 2022). At the same time, significant antibacterial efficiency ( $p < 0.05$ ) was observed in all  
286 modified CNC samples, with >99.999% reduction for both strains. After the chlorination  
287 treatment, MAM-CNCs could generate active chlorine that transferred to cell membranes  
288 and resulted in cell death (Kong et al., 2019). CTAB-CNCs could change the bacterial  
289 membrane permeability or bacterial surface electrostatic balance after contact, ultimately  
290 leading to cell death (Xie et al., 2011). As for ZnO-CNCs, several probable routes have  
291 been proposed to explain their bactericidal mechanism. The released ion mediated killing

292 was one of the possible mechanisms, where the released  $Zn^{2+}$  entered the bacterial cells,  
 293 leading to inhibited DNA replication. ZnO might also attach and induce deformation of the  
 294 bacterial cell wall (Ahmad, 2021; Roy & Rhim, 2019; Zhang et al., 2021; Larki et al., 2022).



295  
 296 **Figure 4.** (a, b) Antibacterial activity and (c, d) images of inhibition zone of PLA/CNC  
 297 composite films (1-Neat PLA, 2-PLA-CC8, 3-PLA-MC8, 4-PLA-ZC8) against *S. aureus*  
 298 and *E. coli*. Means with different letters on the column top were significantly different ( $p$   
 299  $< 0.05$ ).

300 As shown in Figure 4 (a, b), the control groups, including neat PLA film, neat PLA films  
 301 treated with acetate acid, PLA film with pristine CNC coating, and PLA film with MAM-  
 302 CNC coating (without chlorination), did not show any antibacterial activity against either  
 303 *S. aureus* or *E. coli*. On the contrary, all the antibacterial CNC coatings exhibited obvious  
 304 effects, depending on the types of bacteria and CNC modifications, as well as the amount

305 of CNC derivatives. Particularly, PLA-CC composite films displayed an increasing log  
306 reduction of *S. aureus* from 2.14 to 6.29 with increasing CNC contents, while PLA-MC  
307 showed high antibacterial efficiency against *S. aureus* even with low coating mass.  
308 However, the antibacterial activity of PLA-ZC for *S. aureus* was not as high as the other  
309 two coatings. In the case of *E. coli*, all three CNC coatings showed significant antibacterial  
310 efficiencies (>99.9999%) upon 1 hour of contact. It meant that CTAB-CNC and ZnO-CNC  
311 coatings had a stronger influence upon the Gram-negative bacteria than the Gram-positive  
312 strains, and the chlorinated MAM-CNCs showed excellent antibacterial activity for both  
313 types of bacteria. Similar results have reported that *E. coli* was less resistant to the  
314 antibacterial agents (e.g., ZnO nanoparticles) than the Gram-positive bacteria (Shankar et  
315 al., 2018; Zhang et al., 2017). In one aspect, the Gram-positive *S. aureus* consists of a  
316 thicker cell wall than that of the Gram-negative *E. coli*; in another aspect, the Gram-  
317 positive bacteria could form aggregates to protect the internal cells from the antibacterial  
318 agents (Pantani et al., 2013). Based on the antibacterial efficiencies, PLA-CC8, PLA-ZC8,  
319 and PLA-MC8 were selected for the following tests. The inhibition zones of neat and  
320 coated PLA films are shown in Figure 4 (c, d). No obvious inhibition zone was observed  
321 in neat PLA, PLA-CC8, and PLA-MC8 films for both *S. aureus* and *E. coli*, suggesting  
322 little or no release of the antibacterial agents from the coatings to the agar plates. On the  
323 contrary, PLA-ZC8 films exhibited similar inhibition rings (~10 mm) against *S. aureus* and  
324 *E. coli*, which was due to the release of ZnO and/or Zn<sup>2+</sup> ions (Pantani et al., 2013; Wahid  
325 et al., 2019). Table 1 summarizes the antibacterial properties of the recently reported  
326 composite materials, suggesting that the sprayed coatings of modified CNCs provided a  
327 promising strategy for effective bacteria elimination.

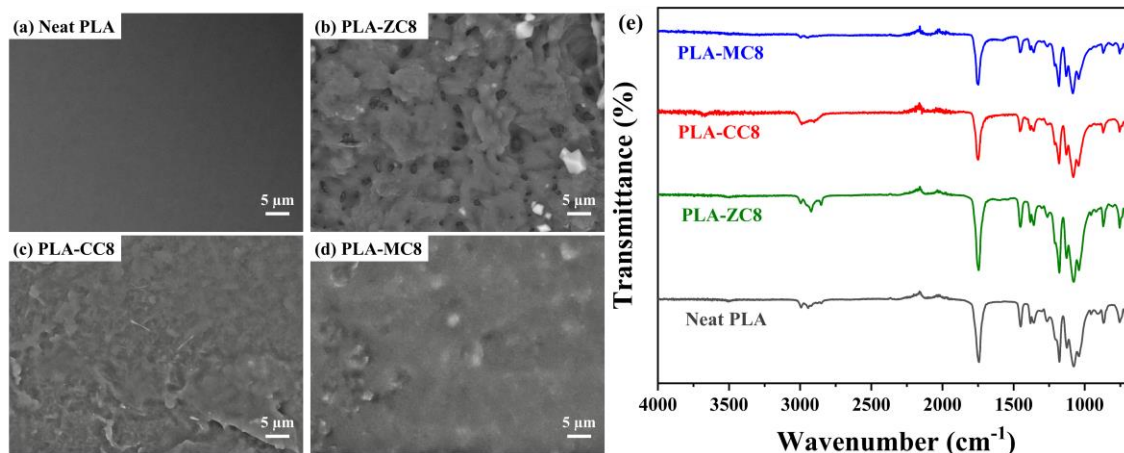
328 **Table 1.** Antibacterial activities of recently reported composite materials.

Composite materials	Antibacterial agent	Antibacterial activity		References
		<i>S. aureus</i>	<i>E. coli</i>	
PLA/CNC nanofluids	CNC nanofluids	98.5%	92.7%	(Shen et al., 2021)
PLA/acetylated CNCs/ZnO	ZnO	99.9%	99.9%	(Yu et al., 2021)
PLA/propolis	Propolis	-	99.99%	(Ulloa et al., 2019)
CMC/gelatin/nano ZnO	CMC & ZnO	84.7%	99.2%	(Chen et al., 2022)
PU/chitin/ZnO-doped-SiO <sub>2</sub> nanoparticles	ZnO-doped-SiO <sub>2</sub> nanoparticles	99.289%	99.942%	(Moustafa et al., 2022)
Graphene/N-halamine-coated cotton fabrics	N-halmaine	99.9999%	99.9999%	(Xu et al., 2022)
Polycaprolactone/Zein/ZnO-QAS	ZnO-QAS nanoparticles	99.9999%	99.9999%	(Wang et al., 2021)
PU/PSDT	N-halmaine & QAS compound	99.9999%	99.9999%	(Tian et al., 2021)
PLA/ZnO-CNCs	ZnO-CNCs	99%	99.9999%	Present work
PLA/CTAB-CNCs	CTAB-CNCs	99.9999%	99.9999%	Present work
PLA/MAM-CNCs	MAM-CNCs	99.9999%	99.9999%	Present work

329 Note: CMC-carboxymethyl chitosan; PU-polyurethane; QAS-quaternary ammonium salts; PSDT-  
 330 polystyrene grafted by 5, 5-dimethylhydantoin and trimethylamine.



331 **3.3. Structure of PLA/CNC composite films**

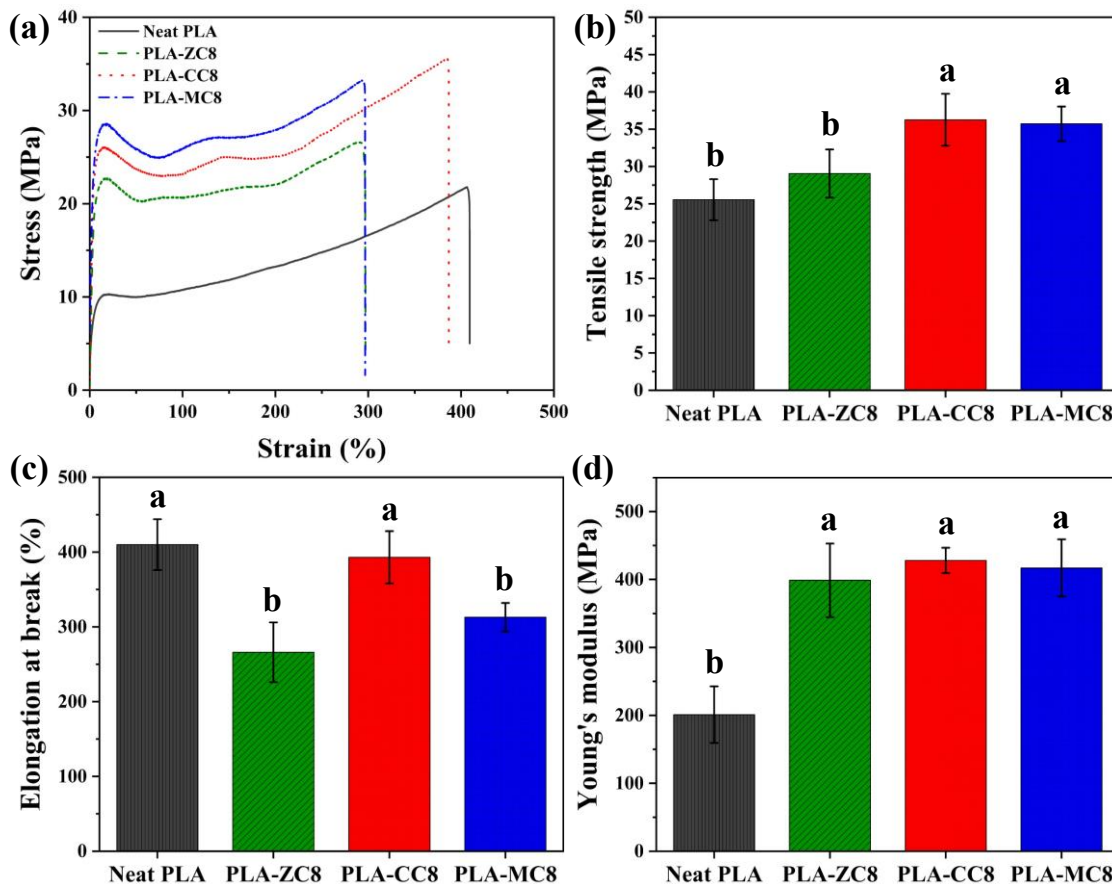


332

333 **Figure 5.** SEM images (a-d) and FTIR spectra (e) of neat and coated PLA films.

334 The surface morphology of neat and coated PLA films was observed by SEM. Figure 5 (a)  
335 displays the smooth and homogeneous surface of neat PLA without cracks or holes. The  
336 addition of coating solutions (acetic acid) resulted in the eroded surfaces, and the modified  
337 CNCs were combined with the polymeric matrix. It explained the little or no release of  
338 CNCs from the films to the agar plates. FTIR analysis of the composite films was carried  
339 out to study the molecular structures. As shown in Figure 5, the peaks of neat PLA film at  
340  $2994\text{ cm}^{-1}$  and  $2943\text{ cm}^{-1}$  were associated with the C-H asymmetric and symmetric  
341 stretching in  $-\text{CH}_3$  groups. The peak at  $1743\text{ cm}^{-1}$  corresponded to the C=O stretching  
342 vibration of the ester groups. The peaks between  $1450\text{ cm}^{-1}$  and  $1357\text{ cm}^{-1}$  represented  
343 asymmetric and symmetric bending of C-H bond in the methyl groups, while the peaks at  
344  $1178\text{ cm}^{-1}$  and  $1077\text{ cm}^{-1}$  were related to the C-O-C symmetric and asymmetric stretching  
345 (Doganay et al., 2016). With the addition of CNC coatings, no new characteristic peaks or  
346 obvious shift of peaks were observed. It indicated that no new covalent bonds were formed  
347 after the incorporation of modified CNCs.

### 3.4. Mechanical and barrier properties of PLA/CNC composite films

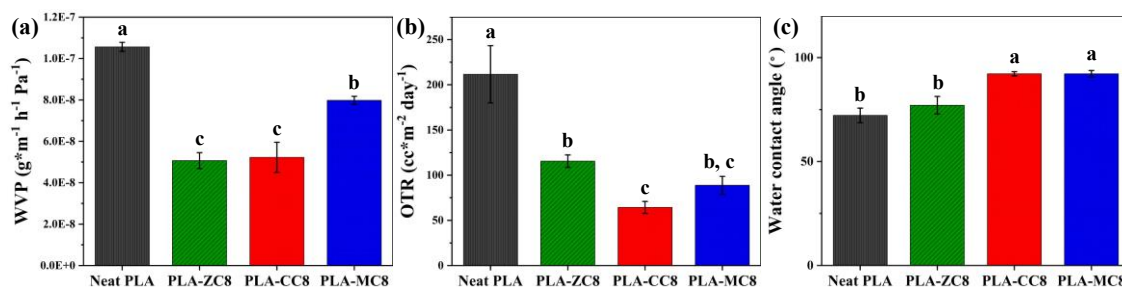


349

350 **Figure 6.** (a) Stress-strain curves, (b) tensile strength, (c) elongation at break, and (d)  
 351 Young's modulus of neat and coated PLA films. Means with different letters on the column  
 352 top were significantly different ( $p < 0.05$ ).

353 Mechanical properties of neat and coated PLA films were measured, and the representative  
 354 stress-strain curves are shown in Figure 6 (a). Their thicknesses did not have any significant  
 355 difference, and all the films exhibited similar curves, beginning with elastic deformation  
 356 and following by plastic deformation. Obvious yielding points were observed, which define  
 357 the limitation of the elastic behavior and the beginning of the plastic behavior (Beauson et  
 358 al., 2022). For the neat PLA films, the stress at the yielding point was ~10 MPa and the  
 359 films fractured at the strain of ~400%. For all types of coated PLA films, an increase in  
 360 yield strength was clearly observed (20-30 MPa), suggesting their improved resistance to  
 361 loading before permanent deformation (Aghajani et al., 2018). Figure 6 (b-d) shows the

362 variations of tensile strength, elongation at break, and Young's modulus of PLA films with  
 363 different CNC coatings. All the coatings significantly increased the Young's modulus, and  
 364 PLA-CC8 and PLA-MC8 had an improved tensile strength, but only PLA-CC8 could  
 365 maintain the ductility of the neat PLA films. The increase in mechanical strength could be  
 366 explained that the surface coating well combined with the polymeric matrix and acted as a  
 367 scaffold (Gulzar et al., 2022). The retained ductility of PLA-CC8 might be attributed to the  
 368 good interfacial compatibility, which contributed to effective stress transfer and delaying  
 369 the stretching failure (Dehnad et al., 2014; Jin et al., 2020).

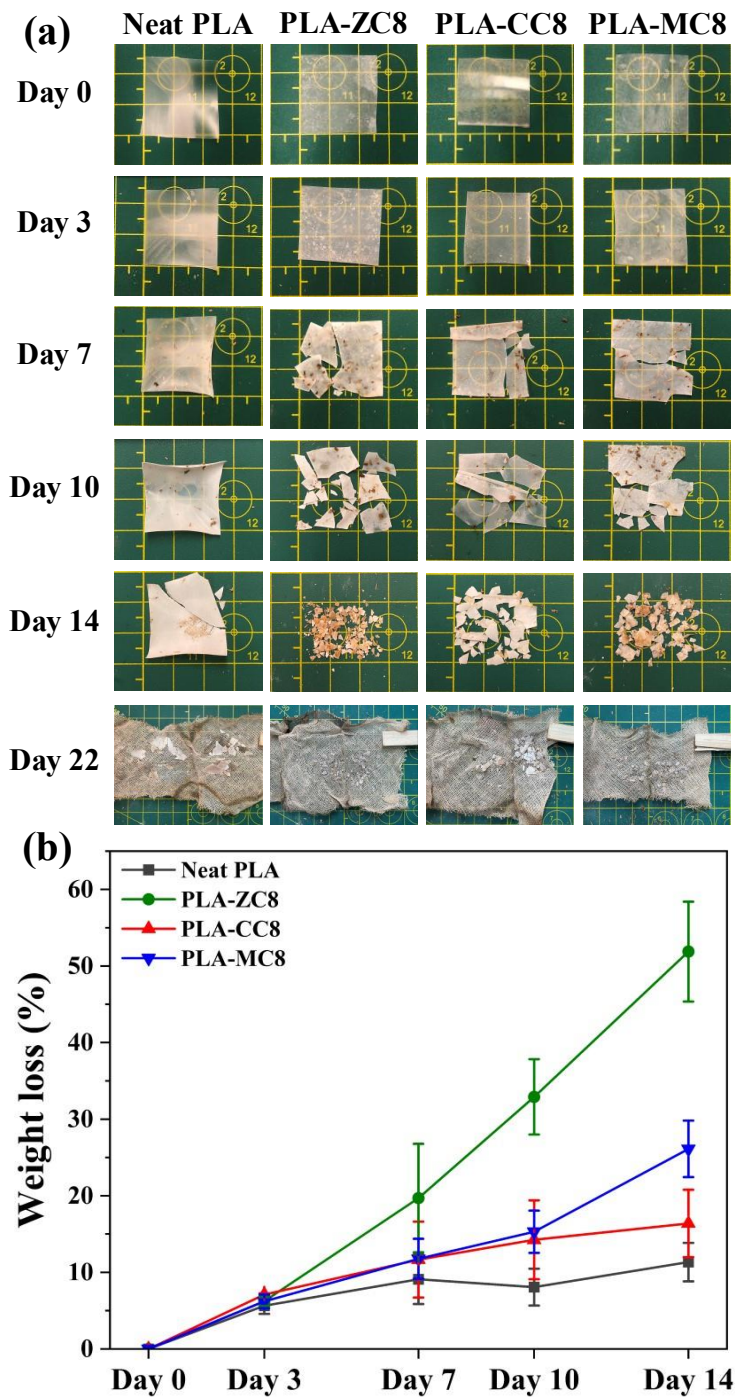


370  
 371 **Figure 7.** (a) WVP, (b) OTR, and (c) water contact angle of neat and coated PLA films.  
 372 Means with different letters on the top of the columns were significantly different ( $p <$   
 373 0.05).

374 The water vapor and oxygen barrier properties of neat and coated PLA films are important  
 375 to their potential applications in packaging. As shown in Figure 7 (a, b), the neat PLA film  
 376 had an average WVP value of  $1.06 \times 10^{-7} \text{ g m}^{-1}\text{h}^{-1}\text{Pa}^{-1}$  and OTR value of  $211.72 \text{ cc m}^{-2}\text{day}^{-1}$ ,  
 377 while the permeability of PLA/CNC composite films significantly ( $p < 0.05$ ) decreased.  
 378 Particularly, PLA-CC8 showed the lowest WVP and OTR values of  $5.22 \times 10^{-8} \text{ g m}^{-1}\text{h}^{-1}\text{Pa}^{-1}$   
 379 and  $64.34 \text{ cc m}^{-2}\text{day}^{-1}$ , respectively. It was because the modified CNCs tightly combined  
 380 with PLA after solution coating and formed a dense layer on the surface of the films (Zhou  
 381 et al., 2021). These values were relatively lower than those reported previously; for  
 382 example, the WVP value of PLA/ZnO composite films was around  $7 \times 10^{-8} \text{ g m}^{-1}\text{h}^{-1}\text{Pa}^{-1}$   
 383 (Shankar et al., 2018), and the poly(ethylene furanoate)/PLA films had an OTR value of  
 384  $144 \text{ cc m}^{-2}\text{day}^{-1}$  (Fredri et al., 2022). Figure 7 (c) shows the water contact angle of neat and  
 385 coated PLA films. As expected, the neat PLA film had a hydrophilic surface with a contact

386 angle of around 74.2° (Vilarinho et al., 2021). A slight increase ( $p > 0.05$ ) in contact angle  
387 of PLA-ZC8 was observed, which might be due to the hydrophobic nature of ZnO (Roy &  
388 Rhim, 2019). The surface hydrophobicity of PLA-CC8 and PLA-MC8 films was  
389 significantly higher than that of the neat PLA film. It could be explained by the existence  
390 of long carbon chains on the modified CNCs (Ly & Mekonnen, 2020).

391 **3.5. Compost disintegrability**



392

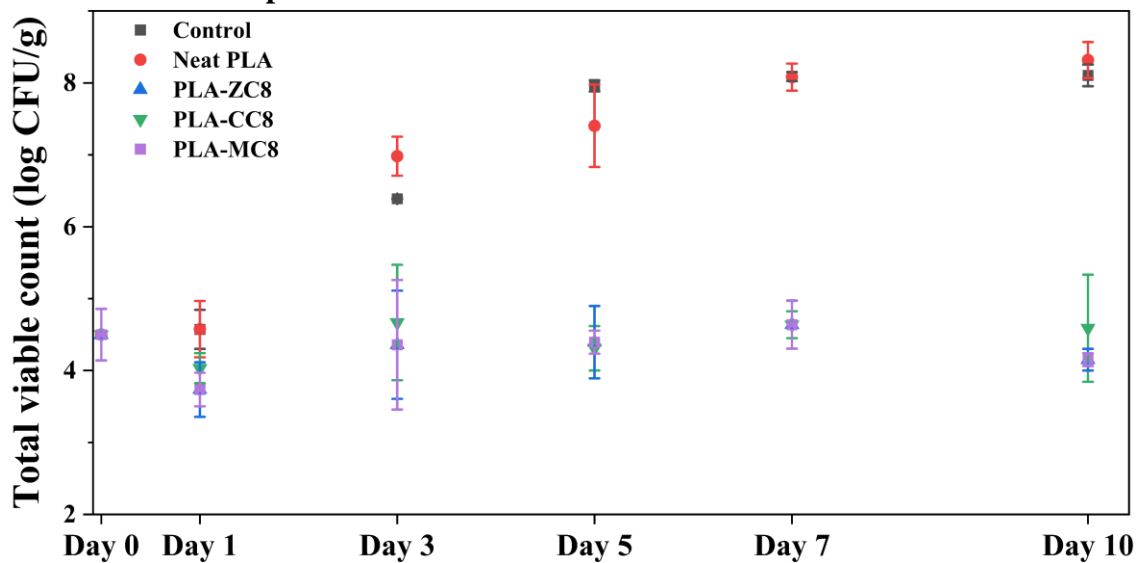
393

394 **Figure 8.** (a) Photos and (b) weight loss of neat and coated PLA films after incubating  
 395 under composting conditions.

396 PLA films hold great potential for “green” packaging materials because of their degradable

397 character under specific conditions (Rojas et al., 2021). In this regard, the effect of CNC  
 398 coatings on the disintegrability of PLA films under composting conditions was investigated.  
 399 As shown in Figure 8 (a), no particular alterations were observed after 3 days for all the  
 400 samples. The PLA/CNC composite films became opaque, and the fragmentation started  
 401 after 7 days. Compared to neat PLA film, the fragmentation of CNC coated PLA films was  
 402 more obvious on day 10 and day 14. The weight loss showed a similar trend (Figure 8 (b)).  
 403 All the films exhibited a gradual increase in weight loss under composting conditions.  
 404 Especially after 14 days, the weight loss of PLA-ZC8, PLA-MC8, and PLA-CC8 films was  
 405 about 51.88%, 26.12%, and 16.38%, respectively, against 11.34% for the neat PLA film.  
 406 To be noted, only a few tiny fragments were recovered on day 22, so it was difficult to  
 407 weigh and calculate the weight loss. The results revealed a faster disintegration rate of  
 408 PLA/CNC composite films, and it was consistent with reported results that CNCs acted as  
 409 a source of energy and carbon and facilitated the initiation of disintegration (Degli-  
 410 Innocenti, 2021; Lizundia et al., 2018; Sun et al., 2022).

### 411 3.6. Preservation of pork



412

413 **Figure 9.** Total viable counts of pork samples during storage at 4 °C.

414 Pork is a commonly consumed red meat worldwide, containing rich sources of certain  
 415 protein, vitamins, and minerals. However, cross-contamination happens frequently during

416 the production. Herein, pork was chosen as a model for evaluating the packaging  
417 performance. The initial TVC value of 4.50 log CFU/g in the raw pork was similar to a  
418 reported value (Vargas Romero et al., 2021). As shown in Figure 9, the total viable  
419 population of the microorganism in pork samples (unpackaged and wrapped with neat PLA  
420 film) rapidly increased during the storage period. The permissible limit of the overall  
421 microorganism in fresh meat should be less than  $5 \times 10^6$  CFU/g, as regulated by European  
422 Commission No. 2073/2005 (Commission, 2005). Therefore, both the control sample and  
423 neat PLA group exceeded the limit after 3 days of cold storage (about 6.39 and 6.98 log  
424 CFU/g). It was worth noting that all PLA/CNC composite film wrapped pork samples had  
425 delayed bacteria growth, and their TVC values were lower than  $5 \times 10^6$  CFU/g after storage  
426 at 4 °C for 10 days. No significant difference was observed among the three coatings, which  
427 indicated the sufficient antibacterial activity of all these modified CNCs for extended shelf  
428 life of fresh pork.

#### 429 **4. Conclusions**

430 This work demonstrated a convenient method to fabricate compostable packaging films  
431 with antibacterial activity. The effects of sprayed coatings of three types of modified CNCs  
432 (CTAB-CNCs, ZnO-CNCs, and MAM-CNCs) on the structure and properties of PLA films  
433 were revealed. Especially, ZnO-CNC coated PLA films exhibited prominent disintegration  
434 behavior. After incubating under composting conditions for 14 days, a weight loss of 51.88%  
435 was recorded, which was over 5 times higher than the neat PLA film. CTAB-CNC coating  
436 significantly increased the tensile strength and Young's modulus without affecting the  
437 ductility, and at the same time, showed the improvement of 51% and 70% in water vapor  
438 and oxygen barrier properties, respectively. MAM-CNC coated PLA films had the most  
439 effective antibacterial capacity against both Gram-positive and Gram-negative bacteria.  
440 With a small amount of coating, PLA-MC4 films could inactivate >99.9999% of *S. aureus*  
441 and *E. coli* upon 1 hour of contact. After all, three PLA/CNC composite films could  
442 significantly extend the shelf life of wrapped pork from 3 days to more than 10 days.  
443 Further investigations are expected to ensure that the biocompatibility inherent to PLA

444 films is retained after coating. Hopefully, this strategy of spray coating with modified  
445 CNCs could be applied in other plastic and paper-based packaging materials.

446 **Author Contributions:** Conceptualization, investigation, and writing, S.H.; Data  
447 collection, S.Z.; writing-review & editing and supervision, Y.W.. All authors have read  
448 and agreed to the published version of the manuscript.

449 **Acknowledgment:** We would like to acknowledge financial support from the Natural  
450 Sciences and Engineering Research Council of Canada (NSERC RGPIN-2019-04498),  
451 Natural Sciences and Engineering Research Council of Canada Discovery Launch  
452 Supplement (NSERC DGEGR-2019-00472), and Canada Foundation for Innovation  
453 (39173). We would like to acknowledge McGill University ECP3 Multi-Scale Imaging  
454 Facility for the image acquisition. S.H. would like to thank the China Scholarship Council  
455 (CSC NO. 201908880002) for financial support of her Ph.D. program.

456 **Conflicts of Interest:** The authors declare no conflict of interest.



457 **5. References**

- 458 Aghajani, M., Wang, M., Cox, L. M., Killgore, J. P., Greenberg, A. R., & Ding, Y. (2018). Influence of  
459 support-layer deformation on the intrinsic resistance of thin film composite membranes. *Journal of*  
460 *Membrane Science*, 567, 49-57. <https://doi.org/10.1016/j.memsci.2018.09.031>
- 461 Ahmad, H. (2021). Celluloses as support materials for antibacterial agents: a review. *Cellulose*, 28(5), 2715-  
462 2761. <https://doi.org/10.1007/s10570-021-03703-2>
- 463 Arnata, I. W., Suprihatin, S., Fahma, F., Richana, N., & Sunarti, T. C. (2020). Cationic modification of  
464 nanocrystalline cellulose from sago fronds. *Cellulose*, 27(6), 3121-3141. [https://doi.org/10.1007/s10570-](https://doi.org/10.1007/s10570-019-02955-3)  
465 019-02955-3
- 466 Asadi, H., Ghalei, S., Handa, H., & Ramasamy, R. P. (2021). Cellulose nanocrystal reinforced silk fibroin  
467 coating for enhanced corrosion protection and biocompatibility of Mg-based alloys for orthopedic  
468 implant applications. *Progress in Organic Coatings*, 161, 106525.  
469 <https://doi.org/10.1016/j.porgcoat.2021.106525>
- 470 Badawy, A. A., Ghanem, A. F., Yassin, M. A., Youssef, A. M., & Rehim, M. H. A. (2021). Utilization and  
471 Characterization of Cellulose Nanocrystals Decorated with Silver and Zinc Oxide Nanoparticles for  
472 Removal of Lead Ion from Wastewater. *Environmental Nanotechnology, Monitoring & Management*, 16,  
473 100501. <https://doi.org/10.1016/j.enmm.2021.100501>
- 474 Baggio, G. M., Camani, P. H., & Rosa, D. S. (2022). Concentration and carbon chain length effects of cationic  
475 surfactant in enzymatic production of cellulose nanostructures. *Journal of Molecular Liquids*, 346,  
476 118231. <https://doi.org/10.1016/j.molliq.2021.118231>
- 477 Beauson, J., Schillani, G., Van der Schueren, L., & Goutianos, S. (2022). The effect of processing conditions  
478 and polymer crystallinity on the mechanical properties of unidirectional self-reinforced PLA composites.  
479 *Composites Part A: Applied Science and Manufacturing*, 152, 106668.  
480 <https://doi.org/10.1016/j.compositesa.2021.106668>
- 481 Cerro, D., Bustos, G., Villegas, C., Buendia, N., Truffa, G., Godoy, M. P., Rodriguez, F., Rojas, A., Galotto,  
482 M.J., Constandil, L., & Torres, A. (2021). Effect of supercritical incorporation of cinnamaldehyde on  
483 physical-chemical properties, disintegration and toxicity studies of PLA/lignin nanocomposites.  
484 *International Journal of Biological Macromolecules*, 167, 255-266.

485 <https://doi.org/10.1016/j.ijbiomac.2020.11.140>

486 Chang, D., Li, Z., Wang, X., Zhu, C., Dong, A., & Gao, G. (2018). N-Halamine polymer from bipolymer to  
487 amphiphilic terpolymer with enhancement in antibacterial activity. *Colloids and Surfaces B:  
488 Biointerfaces*, 163, 402-411. <https://doi.org/10.1016/j.colsurfb.2018.01.013>

489 Chen, J., Luo, L., Cen, C., Liu, Y., Li, H., & Wang, Y. (2022). The nano antibacterial composite film  
490 carboxymethyl chitosan/gelatin/nano ZnO improves the mechanical strength of food packaging.  
491 *International Journal of Biological Macromolecules*, 220, 462-471.  
492 <https://doi.org/10.1016/j.ijbiomac.2022.08.005>

493 Commission Regulation (EC) No 2073/2005 of 15 November 2005 on microbiological criteria for foodstuffs.  
494 (2005). <https://eur-lex.europa.eu/legal-content/EN/ALL/?uri=CELEX%3A32005R2073>

495 Degli-Innocenti, F. (2021). Is composting of packaging real recycling? *Waste Management*, 130, 61-64.  
496 <https://doi.org/10.1016/j.wasman.2021.05.017>

497 Dehnad, D., Emam-Djomeh, Z., Mirzaei, H., Jafari, S.-M., & Dadashi, S. (2014). Optimization of physical  
498 and mechanical properties for chitosan–nanocellulose biocomposites. *Carbohydrate Polymers*, 105, 222-  
499 228. <https://doi.org/10.1016/j.carbpol.2014.01.094>

500 Doganay, D., Coskun, S., Kaynak, C., & Unalan, H. E. (2016). Electrical, mechanical and thermal properties  
501 of aligned silver nanowire/poly lactide nanocomposite films. *Composites Part B: Engineering*, 99, 288-  
502 296. <https://doi.org/10.1016/j.compositesb.2016.06.044>

503 Duarte Urueña, G., Ribeiro, K. C., Prestes, E., Pinheiro, L. A., & Carvalho, B. M. (2021). Extraction of  
504 Cellulose Nanocrystal from Multilayer Packaging Residues Composed of a Mixture of Eucalyptus and  
505 Pine Fibers. *Waste and Biomass Valorization*, 12(10), 5763-5777. [https://doi.org/10.1007/s12649-021-  
506 01383-4](https://doi.org/10.1007/s12649-021-01383-4)

507 Elfeky, A. S., Salem, S. S., Elzaref, A. S., Owda, M. E., Eladawy, H. A., Saeed, A. M., Awad, M.A., Abou-  
508 Zeid, R.E., & Fouda, A. (2020). Multifunctional cellulose nanocrystal /metal oxide hybrid, photo-  
509 degradation, antibacterial and larvicidal activities. *Carbohydrate Polymers*, 230, 115711.  
510 <https://doi.org/10.1016/j.carbpol.2019.115711>

511 Fiore, A., Park, S., Volpe, S., Torrieri, E., & Masi, P. (2021). Active packaging based on PLA and chitosan-  
512 caseinate enriched rosemary essential oil coating for fresh minced chicken breast application. *Food*

513 *Packaging and Shelf Life*, 29, 100708. <https://doi.org/10.1016/j.fpsl.2021.100708>

514 Fonseca-García, A., Jiménez-Regalado, E. J., & Aguirre-Loredo, R. Y. (2021). Preparation of a novel  
515 biodegradable packaging film based on corn starch-chitosan and poloxamers. *Carbohydrate*  
516 *Polymers*, 251, 117009. <https://doi.org/10.1016/j.carbpol.2020.117009>

517 Fredi, G., Dorigato, A., Dussin, A., Xanthopoulou, E., Bikiaris, D.N., Botta, L., Fiore, V. and Pegoretti, A.  
518 (2022). Compatibilization of Polylactide/Poly (ethylene 2, 5-furanoate)(PLA/PEF) Blends for  
519 Sustainable and Bioderived Packaging. *Molecules*, 27(19), 6371.  
520 <https://doi.org/10.3390/molecules27196371>

521 Fu, F., Li, L., Liu, L., Cai, J., Zhang, Y., Zhou, J., & Zhang, L. (2015). Construction of Cellulose Based ZnO  
522 Nanocomposite Films with Antibacterial Properties through One-Step Coagulation. *ACS Applied*  
523 *Materials & Interfaces*, 7(4), 2597-2606. <https://doi.org/10.1021/am507639b>

524 Gahrooee, T. R., Abbasi Moud, A., Danesh, M., & Hatzikiriakos, S. G. (2021). Rheological characterization  
525 of CNC-CTAB network below and above critical micelle concentration (CMC). *Carbohydrate Polymers*,  
526 257, 117552. <https://doi.org/10.1016/j.carbpol.2020.117552>

527 Gao, P., Cha, R., Luo, H., Xu, Y., Zhang, P., Han, L., Wang, X., Zhang, Z., & Jiang, X. (2022). Development  
528 of antimicrobial oxidized cellulose film for active food packaging. *Carbohydrate Polymers*, 278, 118922.  
529 <https://doi.org/10.1016/j.carbpol.2021.118922>

530 Guan, Y., Yu, H.-Y., Abdalkarim, S. Y. H., Wang, C., Tang, F., Marek, J., Chen, W.L., Militky, J., & Yao, J.-  
531 M. (2019). Green one-step synthesis of ZnO/cellulose nanocrystal hybrids with modulated morphologies  
532 and superfast absorption of cationic dyes. *International Journal of Biological Macromolecules*, 132, 51-  
533 62. <https://doi.org/10.1016/j.ijbiomac.2019.03.104>

534 Gulzar, S., Tagrida, M., Nilsuwan, K., Prodpran, T., & Benjakul, S. (2022). Electrospinning of  
535 gelatin/chitosan nanofibers incorporated with tannic acid and chitoooligosaccharides on polylactic acid  
536 film: Characteristics and bioactivities. *Food Hydrocolloids*, 133, 107916.  
537 <https://doi.org/10.1016/j.foodhyd.2022.107916>

538 Huang, S., Liu, X., Chang, C., & Wang, Y. (2020). Recent developments and prospective food-related  
539 applications of cellulose nanocrystals: a review. *Cellulose*, 27(6), 2991-3011.  
540 <https://doi.org/10.1007/s10570-020-02984-3>

541 Huang, S., Tao, R., Ismail, A., & Wang, Y. (2020). Cellulose Nanocrystals Derived from Textile Waste  
542 through Acid Hydrolysis and Oxidation as Reinforcing Agent of Soy Protein Film. *Polymers*, 12(4), 958.  
543 <https://doi.org/10.3390/polym12040958>

544 International Organization for Standardization. (2015). *Plastics-Determination of the degree of disintegration*  
545 *of plastic materials under simulated composting conditions in a laboratory-scale test* (ISO 20200:2015).  
546 <https://www.iso.org/standard/63367.html>

547 Jiang, S., Li, Q., Wang, F., Wang, Z., Cao, X., Shen, X., & Yao, Z. (2022). Highly effective and sustainable  
548 antibacterial membranes synthesized using biodegradable polymers. *Chemosphere*, 291, 133106.  
549 <https://doi.org/10.1016/j.chemosphere.2021.133106>

550 Jin, K., Tang, Y., Zhu, X., & Zhou, Y. (2020). Polylactic acid based biocomposite films reinforced with  
551 silanized nanocrystalline cellulose. *International Journal of Biological Macromolecules*, 162, 1109-1117.  
552 <https://doi.org/10.1016/j.ijbiomac.2020.06.201>

553 Kong, X., Zhang, S., Wang, Y., Liu, Y., Li, R., Ren, X., & Huang, T.-S. (2019). Antibacterial polyvinyl alcohol  
554 films incorporated with N-halamine grafted oxidized microcrystalline cellulose. *Composites*  
555 *Communications*, 15, 25-29. <https://doi.org/10.1016/j.coco.2019.06.005>

556 Kongkaoroptham, P., Piroonpan, T., & Pasanphan, W. (2021). Chitosan nanoparticles based on their  
557 derivatives as antioxidant and antibacterial additives for active bioplastic packaging. *Carbohydrate*  
558 *Polymers*, 257, 117610. <https://doi.org/10.1016/j.carbpol.2020.117610>

559 Koshani, R., Zhang, J., van de Ven, T. G. M., Lu, X., & Wang, Y. (2021). Modified Hairy Nanocrystalline  
560 Cellulose as Photobactericidal Nanofillers for Food Packaging Application. *ACS Sustainable Chemistry*  
561 *& Engineering*, 9(31), 10513-10523. <https://doi.org/10.1021/acssuschemeng.1c02289>

562 Larki, M., Enayati, M. H., & Rostamabadi, H. (2022). Basil seed gum promotes the electrospinnability of  
563 WPI for co-encapsulation of ZnO nanoparticles and curcumin. *Carbohydrate Polymers*, 296, 119966.  
564 <https://doi.org/10.1016/j.carbpol.2022.119966>

565 Li, M.-C., Wu, Q., Han, J., Mei, C., Lei, T., Lee, S.-y., & Gwon, J. (2020). Overcoming Salt Contamination  
566 of Bentonite Water-Based Drilling Fluids with Blended Dual-Functionalized Cellulose Nanocrystals.  
567 *ACS Sustainable Chemistry & Engineering*, 8(31), 11569-11578.  
568 <https://doi.org/10.1021/acssuschemeng.0c02774>

569 Liu, Y., Li, L., Pan, N., Wang, Y., Ren, X., Xie, Z., Buschle-Diller, G., & Huang, T.-S. (2017). Antibacterial  
570 cellulose acetate films incorporated with N-halamine-modified nano-crystalline cellulose particles.  
571 *Polymers for Advanced Technologies*, 28(4), 463-469. <https://doi.org/10.1002/pat.3906>

572 Lizundia, E., Goikuria, U., Vilas, J. L., Cristofaro, F., Bruni, G., Fortunati, E., Armentano, I., Visai, L., &  
573 Torre, L. (2018). Metal Nanoparticles Embedded in Cellulose Nanocrystal Based Films: Material  
574 Properties and Post-use Analysis. *Biomacromolecules*, 19(7), 2618-2628.  
575 <https://doi.org/10.1021/acs.biomac.8b00243>

576 Ly, M., & Mekonnen, T. H. (2020). Cationic surfactant modified cellulose nanocrystals for corrosion  
577 protective nanocomposite surface coatings. *Journal of Industrial and Engineering Chemistry*, 83, 409-  
578 420. <https://doi.org/10.1016/j.jiec.2019.12.014>

579 Ma, X., Wu, N., Liu, P., & Cui, H. (2022). Fabrication of highly efficient phenylphosphorylated chitosan bio-  
580 based flame retardants for flammable PLA biomaterial. *Carbohydrate Polymers*, 287, 119317.  
581 <https://doi.org/10.1016/j.carbpol.2022.119317>

582 Moustafa, H., Darwish, N. A., & Youssef, A. M. (2022). Rational formulations of sustainable  
583 polyurethane/chitin/rosin composites reinforced with ZnO-doped-SiO<sub>2</sub> nanoparticles for green packaging  
584 applications. *Food Chemistry*, 371, 131193. <https://doi.org/10.1016/j.foodchem.2021.131193>

585 Niinivaara, E., Desmaisons, J., Dufresne, A., Bras, J., & Cranston, E. D. (2021). Thick Polyvinyl Alcohol  
586 Films Reinforced with Cellulose Nanocrystals for Coating Applications. *ACS Applied Nano Materials*.  
587 <https://doi.org/10.1021/acsanm.1c01244>

588 Omerović, N., Djisalov, M., Živojević, K., Mladenović, M., Vunduk, J., Milenković, I., Gadjanski, I., & Vidić,  
589 J. (2021). Antimicrobial nanoparticles and biodegradable polymer composites for active food packaging  
590 applications. *Comprehensive Reviews in Food Science and Food Safety*, 20(3), 2428-2454.  
591 <https://doi.org/10.1111/1541-4337.12727>

592 Pantani, R., Gorrasi, G., Vigliotta, G., Murariu, M., & Dubois, P. (2013). PLA-ZnO nanocomposite films:  
593 Water vapor barrier properties and specific end-use characteristics. *European Polymer Journal*, 49(11),  
594 3471-3482. <https://doi.org/10.1016/j.eurpolymj.2013.08.005>

595 Park, S., Baker, J. O., Himmel, M. E., Parilla, P. A., & Johnson, D. K. (2010). Cellulose crystallinity index:  
596 measurement techniques and their impact on interpreting cellulase performance. *Biotechnol Biofuels*, 3(1),

597 10. <https://doi.org/10.1186/1754-6834-3-10>

598 Ranjbar, D., Raeiszadeh, M., Lewis, L., MacLachlan, M. J., & Hatzikiriakos, S. G. (2020). Adsorptive  
599 removal of Congo red by surfactant modified cellulose nanocrystals: a kinetic, equilibrium, and  
600 mechanistic investigation. *Cellulose*, 27(6), 3211-3232. <https://doi.org/10.1007/s10570-020-03021-z>

601 Rojas-Lema, S., Nilsson, K., Trifol, J., Langton, M., Gomez-Caturla, J., Balart, R., Garcia-Garcia, D. and  
602 Moriana, R. (2021). Faba bean protein films reinforced with cellulose nanocrystals as edible food  
603 packaging material. *Food Hydrocolloids*, 121, 107019. <https://doi.org/10.1016/j.foodhyd.2021.107019>

604 Rojas, A., Velásquez, E., Patiño Vidal, C., Guarda, A., Galotto, M. J., & López de Dicastillo, C. (2021). Active  
605 PLA Packaging Films: Effect of Processing and the Addition of Natural Antimicrobials and Antioxidants  
606 on Physical Properties, Release Kinetics, and Compostability. *Antioxidants*, 10(12), 1976.  
607 <https://doi.org/10.3390/antiox10121976>

608 Rosace, G., Colleoni, C., Trovato, V., Iacono, G., & Malucelli, G. (2017). Vinylphosphonic  
609 acid/methacrylamide system as a durable intumescent flame retardant for cotton fabric. *Cellulose*, 24(7),  
610 3095-3108. <https://doi.org/10.1007/s10570-017-1294-x>

611 Roy, S., & Rhim, J.-W. (2019). Carrageenan-based antimicrobial bionanocomposite films incorporated with  
612 ZnO nanoparticles stabilized by melanin. *Food Hydrocolloids*, 90, 500-507.  
613 <https://doi.org/10.1016/j.foodhyd.2018.12.056>

614 Salmieri, S., Islam, F., Khan, R.A., Hossain, F.M., Ibrahim, H.M., Miao, C., Hamad, W.Y. and Lacroix, M.  
615 (2014a). Antimicrobial nanocomposite films made of poly(lactic acid)-cellulose nanocrystals (PLA-CNC)  
616 in food applications: part A—effect of nisin release on the inactivation of *Listeria monocytogenes* in ham.  
617 *Cellulose*, 21(3), 1837-1850. <https://doi.org/10.1007/s10570-014-0230-6>

618 Salmieri, S., Islam, F., Khan, R.A., Hossain, F.M., Ibrahim, H.M., Miao, C., Hamad, W.Y. and Lacroix, M.  
619 (2014b). Antimicrobial nanocomposite films made of poly(lactic acid)-cellulose nanocrystals (PLA-  
620 CNC) in food applications—part B: effect of oregano essential oil release on the inactivation of *Listeria*  
621 *monocytogenes* in mixed vegetables. *Cellulose*, 21(6), 4271-4285. [https://doi.org/10.1007/s10570-014-](https://doi.org/10.1007/s10570-014-0406-0)  
622 0406-0

623 Shankar, S., Wang, L.-F., & Rhim, J.-W. (2018). Incorporation of zinc oxide nanoparticles improved the  
624 mechanical, water vapor barrier, UV-light barrier, and antibacterial properties of PLA-based

625 nanocomposite films. *Materials Science and Engineering: C*, *93*, 289-298.  
626 <https://doi.org/10.1016/j.msec.2018.08.002>

627 Sharafi Zamir, S., Fathi, B., Ajjji, A., Robert, M., & Elkoun, S. (2022). Crystallinity and Gas Permeability of  
628 Poly (Lactic Acid)/Starch Nanocrystal Nanocomposite. *Polymers*, *14*(14), 2802.  
629 <https://doi.org/10.3390/polym14142802>

630 Sharma, P. R., Sharma, S. K., Antoine, R., & Hsiao, B. S. (2019). Efficient Removal of Arsenic Using Zinc  
631 Oxide Nanocrystal-Decorated Regenerated Microfibrillated Cellulose Scaffolds. *ACS Sustainable*  
632 *Chemistry & Engineering*, *7*(6), 6140-6151. <https://doi.org/10.1021/acssuschemeng.8b06356>

633 Shen, H., Li, Y., Yao, W., Yang, S., Yang, L., Pan, F., Chen, Z., & Yin, X. (2021). Solvent-free cellulose  
634 nanocrystal fluids for simultaneous enhancement of mechanical properties, thermal conductivity,  
635 moisture permeability and antibacterial properties of polylactic acid fibrous membrane. *Composites Part*  
636 *B: Engineering*, *222*, 109042. <https://doi.org/10.1016/j.compositesb.2021.109042>

637 Shin, H., Kim, S., Kim, J., Kong, S., Lee, Y., & Lee, J.-C. (2022). Preparation of 3-pentadecylphenol-  
638 modified cellulose nanocrystal and its application as a filler to polypropylene nanocomposites having  
639 improved antibacterial and mechanical properties. *Journal of Applied Polymer Science*, *139*(13), 51848.  
640 <https://doi.org/10.1002/app.51848>

641 Shojaeiarani, J., Bajwa, D. S., Stark, N. M., Bergholz, T. M., & Kraft, A. L. (2020). Spin coating method  
642 improved the performance characteristics of films obtained from poly(lactic acid) and cellulose  
643 nanocrystals. *Sustainable Materials and Technologies*, *26*, e00212.  
644 <https://doi.org/10.1016/j.susmat.2020.e00212>

645 Shojaeiarani, J., Shirzadifar, A., Shine, C., & Reisi, A. M. (2022). Hybrid nanocomposite packaging films  
646 from cellulose nanocrystals, zinc sulfide quantum dots reinforced polylactic acid with fluorescent and  
647 antibacterial properties. *Polymer Engineering & Science*, *62*, 1562-  
648 1570. <https://doi.org/10.1002/pen.25944>

649 Sun, C., Li, C., Li, H., Liu, M., Zheng, H., Tan, H., Wang, Y., & Zhang, Y. (2022). Modified Cellulose  
650 Nanocrystals Enhanced the Compatibility Between PLA and PBAT to Prepare a Multifunctional  
651 Composite Film. *Journal of Polymers and the Environment*, *30*, 3139–3149.  
652 <https://doi.org/10.1007/s10924-022-02422-4>

653 Tang, X., Xu, H., Shi, Y., Wu, M., Tian, H., & Liang, J. (2020). Porous antimicrobial starch particles  
654 containing N-halamine functional groups. *Carbohydrate Polymers*, 229, 115546.  
655 <https://doi.org/10.1016/j.carbpol.2019.115546>

656 Tian, C., Wu, F., Jiao, W., Liu, X., Yin, X., Si, Y., Yu, J., & Ding, B. (2021). Antibacterial and antiviral N-  
657 halamine nanofibrous membranes with nanonet structure for bioprotective applications. *Composites*  
658 *Communications*, 24, 100668. <https://doi.org/10.1016/j.coco.2021.100668>

659 Ulloa, P. A., Vidal, J., Lopéz de Dicastillo, C., Rodriguez, F., Guarda, A., Cruz, R. M. S., & Galotto, M. J.  
660 (2019). Development of poly(lactic acid) films with propolis as a source of active compounds:  
661 Biodegradability, physical, and functional properties. *Journal of Applied Polymer Science*, 136(8), 47090.  
662 <https://doi.org/10.1002/app.47090>

663 Vargas Romero, E., Lim, L. T., Suarez Mahecha, H., & Bohrer, B. M. (2021). The Effect of Electrospun  
664 Polycaprolactone Nonwovens Containing Chitosan and Propolis Extracts on Fresh Pork Packaged in  
665 Linear Low-Density Polyethylene Films. *Foods*, 10(5), 1110. <https://doi.org/10.3390/foods10051110>

666 Vilarinho, F., Stanzione, M., Buonocore, G. G., Barbosa-Pereira, L., Sendón, R., Vaz, M. F., & Sanches Silva,  
667 A. (2021). Green tea extract and nanocellulose embedded into polylactic acid film: Properties and  
668 efficiency on retarding the lipid oxidation of a model fatty food. *Food Packaging and Shelf Life*, 27,  
669 100609. <https://doi.org/10.1016/j.fpsl.2020.100609>

670 Wahid, F., Duan, Y. X., Hu, X. H., Chu, L. Q., Jia, S. R., Cui, J. D., & Zhong, C. (2019). A facile construction  
671 of bacterial cellulose/ZnO nanocomposite films and their photocatalytic and antibacterial properties.  
672 *International Journal of Biological Macromolecules*, 132, 692-700.  
673 <https://doi.org/10.1016/j.ijbiomac.2019.03.240>

674 Wang, Y., Ying, M., Zhang, M., Ren, X., & Kim, I. S. (2021). Development of Antibacterial and Hemostatic  
675 PCL/Zein/ZnO-Quaternary Ammonium Salts NPs Composite Mats as Wound Dressings.  
676 *Macromolecular Materials and Engineering*, 306(12), 2100587.  
677 <https://doi.org/10.1002/mame.202100587>

678 World Health Organization. (2020). Food safety. World Health Organization. Retrieved August 31, 2022,  
679 from <https://www.who.int/news-room/fact-sheets/detail/food-safety>

680 Xiao, Y., Liu, Y., Kang, S., Wang, K., & Xu, H. (2020). Development and evaluation of soy protein isolate-



681 based antibacterial nanocomposite films containing cellulose nanocrystals and zinc oxide nanoparticles.  
682 *Food Hydrocolloids*, 106, 105898. <https://doi.org/10.1016/j.foodhyd.2020.105898>

683 Xie, D., Weng, Y., Guo, X., Zhao, J., Gregory, R. L., & Zheng, C. (2011). Preparation and evaluation of a  
684 novel glass-ionomer cement with antibacterial functions. *Dental Materials*, 27(5), 487-496.  
685 <https://doi.org/10.1016/j.dental.2011.02.006>

686 Xu, D., Wang, S., Hu, J., Liu, Y., Jiang, Z., & Zhu, P. (2021). Enhancing antibacterial and flame-retardant  
687 performance of cotton fabric with an iminodiacetic acid-containing N-halamine. *Cellulose*, 28(5), 3265-  
688 3277. <https://doi.org/10.1007/s10570-021-03716-x>

689 Xu, Z., Pan, N., Xue, Y., Jiang, T., & Wang, Y. (2022). Long-lasting renewable antibacterial graphene/N-  
690 halamine-coated cotton fabrics benefitting from enhanced UV stability and quantitative tracking of  
691 bactericidal factors. *International Journal of Biological Macromolecules*, 222, 1192-1200.  
692 <https://doi.org/10.1016/j.ijbiomac.2022.09.254>

693 Yu, F., Fei, X., He, Y., & Li, H. (2021). Poly(lactic acid)-based composite film reinforced with acetylated  
694 cellulose nanocrystals and ZnO nanoparticles for active food packaging. *International Journal of*  
695 *Biological Macromolecules*, 186, 770-779. <https://doi.org/10.1016/j.ijbiomac.2021.07.097>

696 Zhang, H., Hortal, M., Jordá-Beneyto, M., Rosa, E., Lara-Lledo, M., & Lorente, I. (2017). ZnO-PLA  
697 nanocomposite coated paper for antimicrobial packaging application. *LWT*, 78, 250-257.  
698 <https://doi.org/10.1016/j.lwt.2016.12.024>

699 Zhang, R., Lan, W., Ji, T., Sameen, D. E., Ahmed, S., Qin, W., & Liu, Y. (2021). Development of polylactic  
700 acid/ZnO composite membranes prepared by ultrasonication and electrospinning for food packaging.  
701 *LWT*, 135, 110072. <https://doi.org/10.1016/j.lwt.2020.110072>

702 Zhang, X., Zhang, Q., Xue, Y., Wang, Y., Zhou, X., Li, Z., & Li, Q. (2021). Simple and green synthesis of  
703 calcium alginate/AgCl nanocomposites with low-smoke flame-retardant and antimicrobial properties.  
704 *Cellulose*, 28(8), 5151-5167. <https://doi.org/10.1007/s10570-021-03825-7>

705 Zhao, S.-W., Zheng, M., Zou, X.-H., Guo, Y., & Pan, Q.-J. (2017). Self-Assembly of Hierarchically  
706 Structured Cellulose@ZnO Composite in Solid-Liquid Homogeneous Phase: Synthesis, DFT  
707 Calculations, and Enhanced Antibacterial Activities. *ACS Sustainable Chemistry & Engineering*, 5(8),  
708 6585-6596. <https://doi.org/10.1021/acs.suschemeng.7b00842>

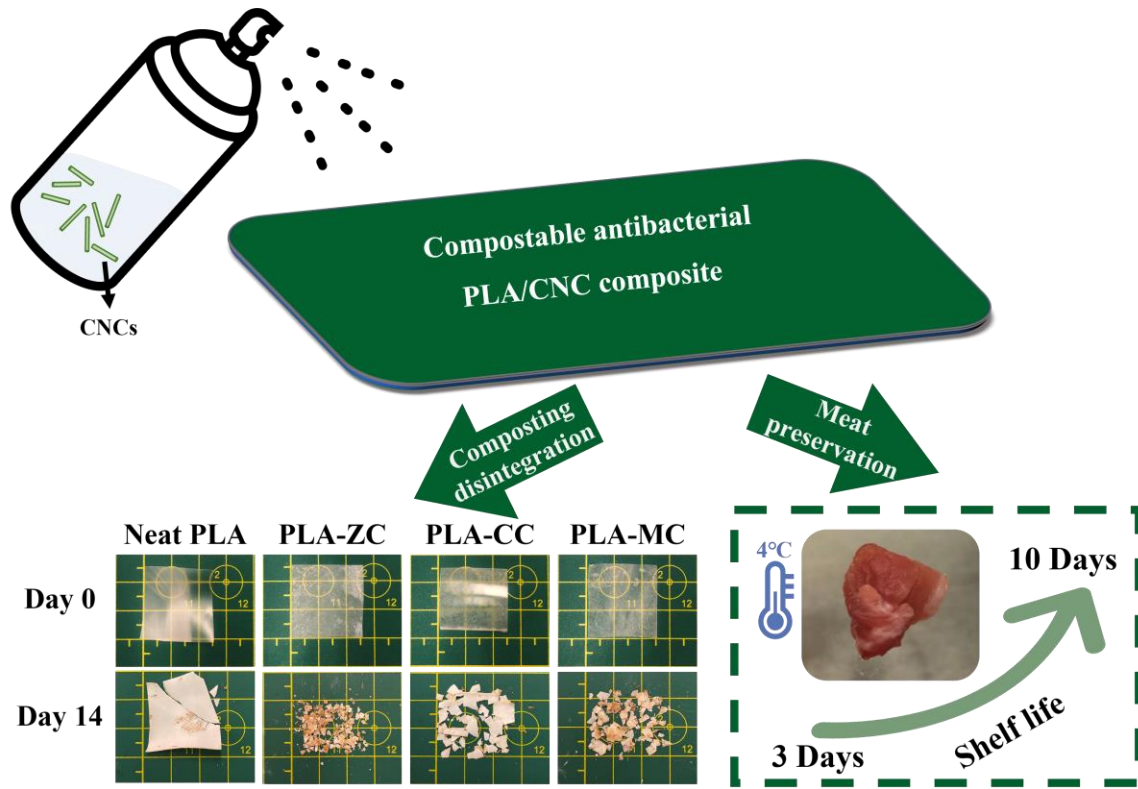
709 Zhong, C., Hou, P.-F., Li, Y.-X., Yang, W.-Y., Shu, M., & Wu, G.-P. (2021). Characterization, antioxidant and  
710 antibacterial activities of gelatin film incorporated with protocatechuic acid and its application on beef  
711 preservation. *LWT*, *151*, 112154. <https://doi.org/10.1016/j.lwt.2021.112154>

712 Zhou, C., Girouard, F., O'Brien, B., Ronholm, J., & Wang, Y. (2022). Construction of chevaux-de-frise from  
713 cellulose nanocrystals to enable mechano-bactericidal activity on recycled waste cotton films. *Green*  
714 *Chemistry*, *24*(3), 1109-1113. <https://doi.org/10.1039/d2gc00073c>

715 Zhou, Q., Chen, J., Wang, C., Yang, G., Janaswamy, S., Xu, F., & Liu, Z. (2022). Preparation and  
716 characterization of lignin nanoparticles and chitin nanofibers reinforced PVA films with UV shielding  
717 properties. *Industrial Crops and Products*, *188*, 115669. <https://doi.org/10.1016/j.indcrop.2022.115669>

718 Zhou, X., Cheng, R., Wang, B., Zeng, J., Xu, J., Li, J., Kang, L., Cheng, Z., Gao, W., & Chen, K. (2021).  
719 Biodegradable sandwich-architected films derived from pea starch and polylactic acid with enhanced  
720 shelf-life for fruit preservation. *Carbohydrate Polymers*, *251*, 117117.  
721 <https://doi.org/10.1016/j.carbpol.2020.117117>

722 Graphical abstract



723